Institutional Biosafety Committee Meeting Minutes

La Jolla Institute FOR IMMUNOLOGY | Life Without Disease.

October 8th, 2025 meeting

A regular meeting of the Institutional Biosafety Committee of the La Jolla Institute for Immunology was held in person on Wednesday, October 8th, 2025, at 9:30 AM, with the option to join via Zoom teleconference.

The meeting started at 9:33 AM.

IBC ATTENDANCE: 11 MEMBERS (9 VOTING MEMBERS, 7 MEMBERS REQUIRED FOR QUORUM)

Regular Members	Present
Miguel Reina-Campos, Ph.D. (Chair)	☑
Mike Barajas (Alternate) CMAR, RLATG	☑
Sylvie Blondelle, Ph.D.	☐
Laurence Cagnon, Ph.D.	☑
Beth Ford, D.V.M.	☑
David Hall, CSP	☑
Peter Jones, BS, LATG	☑
Alessandro Sette, Ph.D.	☐
Stephen Schoenberger, Ph.D.	☐
Kristine Suchey, BS, RVT	☑
Renna Wolfe, Ph.D.	☑
Jeremy Young, BS, MBA	☐
Marianne Zupanc, Ph.D.	☑
Others Present: Jason Vo, Hayley Simon	

Note: Marianne Zupanc joined via zoom.

Laurence Cagnon will be the new vice-chair

REVIEW AND APPROVAL OF THE MINUTES

The August meeting minutes were unanimously approved.

PROTOCOL REVIEW

The risk assessment evaluation matrix was reviewed with the IBC before protocols review. The following terms and percentages reflect the likelihood of an exposure:

- Very significant (>10% risk)
- Significant (1-10% risk)
- Unlikely (0.1-0.99% risk)
- Very unlikely (<0.01% risk)

NEW PROTOCOLS

PI	**Kronenberg**
Protocol #	**BHR30-MK**
Title	Acinetobacter baumannii
Experimental Procedures	
Agent	Acinetobacter baumannii (strain 5377)
Project summary (from form)	Mucosal-associated invariant T cells (MAIT cells) are a specialized immune cell responsible against several types of bacteria. Acinetobacter baumanniii is a bacteria commonly found in water and soil, and infections with this bacteria are increasingly happening in hospital settings, due to the emergence of antibiotic multi-resistant strains. It has been shown that this bacteria can induce strong lung MAIT cells responses that differ from the responses generated from other pathogenic bacteria. Our overall goal is to determine if infections with Acinetobacter baumannii can induce long term changes in these cells that increase protection to future infections.
Additional details from the protocol	N/A
Manipulations planned	Bacteria culture, centrifugation, transport, in vivo infection (injections)
Recombinant or Synthetic Nucleic Acids	
Source of nucleic sequences (e.g., species)	N/A
Nature of nucleic acid (NA) sequences (e.g., enzyme, oncogene)	N/A
NA Host(s) and Vector(s)	N/A
Risk Assessment/Training	
Proposed Risk Assessment	Low
Training	Verified and on record
IBC Assessment	
Proposed Biosafety Level	BSL-2 and ABSL-2
CA ATP-L	No
NIH Guidelines	No
Category 1 Research	No
Category 2 Research	No
IBC Approval	
Unanimously approved at the proposed biosafety levels with the discussed modifications	

PI	**Peters**
Protocol #	**BHR08-BP**
Title	Heat inactivated bacteria
Experimental Procedures	
Agents	Heat inactivated bacteria: - Staphylococcus aureus USA 300 strain and Streptococcus pneumoniae
Project summary (from form)	Using different inactivated bacteria, we will study the immune response in human PBMCs.
Additional details from the protocol	N/A
Manipulations planned	Cell culture, Fluorospot, Flow cytometry with human PBMCs stimulated by heat inactivated bacteria
Recombinant or Synthetic Nucleic Acids	
Source of nucleic sequences (e.g., species)	N/A

Nature of nucleic acid (NA) sequences (e.g., enzyme, oncogene)	N/A
NA Host(s) and Vector(s)	N/A
Risk Assessment/Training	
Proposed Risk Assessment	Low
Training	Verified and on record
IBC Assessment	
Proposed Biosafety Level	BSL-1 (agent alone), BSL-2 (agent on human PBMC)
CA ATP-L	No
NIH Guidelines	No
Category 1 Research	No
Category 2 Research	No
IBC Approval	
Unanimously approved at the proposed biosafety levels	

PI	**Saphire**
Protocol #	**BHR17-ES**
Title	Mumps virus
Experimental Procedures	
Agent	Mumps Virus (Genotype A or G)
Project summary (from form)	I will directly visualize and functionally analyze the human polyclonal and monoclonal antibody response to mumps virus (MuV) infection and vaccination. As there are recurring breakthrough infections with MuV mainly in fully vaccinated populations, we want to illuminate why these breakthrough infections occur and if a lack of cross-reactivity between the circulating and the vaccine strain is the reason for these infections. We will purify antibodies from memory B cells contained in blood samples from MMR recipients. We will then screen antibodies for their ability to bind to MuV proteins and for their ability to neutralize MuV infection in vitro. For antibodies of interest, we will use cryo-electron microscopy to directly map antibody binding sites on MuV proteins. This information will help us learn which sites on MuV proteins are important for viral neutralization and guide us in the design of novel therapeutics and more guided vaccine design.
Additional details from the protocol	Mumps has an R0 of 10-12
Manipulations planned	Tissue culture, viral production, in vitro neutralization assays, centrifugation in aerosol containing devices.
Recombinant or Synthetic Nucleic Acids	
Source of nucleic sequences (e.g., species)	N/A
Nature of nucleic acid (NA) sequences (e.g., enzyme, oncogene)	N/A
NA Host(s) and Vector(s)	N/A
Risk Assessment/Training	
Proposed Risk Assessment	Medium
Training	Verified and on record
IBC Assessment	
Proposed Biosafety Level	BSL-2 with BSL-3 practices aimed at containing the aerosols
CA ATP-L	Yes
NIH Guidelines	N/A (the lab will submit an amendment if using a recombinant virus)
Category 1 Research	N/A

Category 2 Research	N/A
IBC Approval	
Unanimously approved at the proposed biosafety level with the discussed modifications	

PI	**Tawani**
Protocol #	**BHR01-AT**
Title	Xenograft model for testing the efficacy of ADC and CART cells
Experimental Procedures	
Agent	Human cancer cell lines from ATCC: hepG2, Hep3B, D425, Raji, Daudi At this time CART cells will not be used, they will be added at a later time.
Project summary (from form)	We want to test the efficacy of antibody drug conjugates (ADC) against cell lines derived xenograft models.
Additional details from the protocol	N/A
Manipulations planned	In vivo injections
Recombinant or Synthetic Nucleic Acids	
Source of nucleic sequences (e.g., species)	N/A
Nature of nucleic acid (NA) sequences (e.g., enzyme, oncogene)	N/A
NA Host(s) and Vector(s)	N/A
Risk Assessment/Training	
Proposed Risk Assessment	Low
Training	Verified and on record
IBC Assessment	
Proposed Biosafety Level	ABSL-1
CA ATP-L	No
NIH Guidelines	No
Category 1 Research	No
Category 2 Research	No
IBC Approval	
10/08/2025 IBC Meeting: Not approved, more information and clarifications were requested by the IBC. 10/30/2025 IBC Meeting: Unanimously approved at the proposed biosafety level. **Note:** A second IBC meeting was conducted on 10/30/2025, to review BHR01-AT again, after the laboratory provided supplemental explanations. The protocol was modified to reflect the new information. The 8 voting attendees were: Laurence Cagnon, Beth Ford (Via zoom), David Hall, Peter Jones, Stephen Schoenberger, Kristine Suchey, Marianne Zupanc) and Mike Barajas. The meeting was convened in person and via zoom, quorum was reached.	

RENEWALS

PI	**Myers**
Protocol #	**BHR01-SM**
Title	Myers - Human Blood
Experimental Procedures	
Agent	Normal Human Blood
Project summary (from form)	Isolate and genetically modify human immune cells to understand signaling and gene expression pathways.

Additional details from the protocol	Note: Primary human T cells will be isolated from blood samples collected through the LJI normal donor program
Manipulations planned	PBMC purification from whole blood, tissue culture, in vitro assays
Recombinant or Synthetic Nucleic Acids	
Source of nucleic sequences (e.g., species)	N/A
Nature of nucleic acid (NA) sequences (e.g., enzyme, oncogene)	N/A
NA Host(s) and Vector(s)	N/A
Risk Assessment/Training	
Proposed Risk Assessment	Low
Training	Verified and on record
IBC Assessment	
Assigned Biosafety Level	BSL-2
CA ATP-L	No
NIH Guidelines	N/A
Category 1 Research	No
Category 2 Research	No
IBC Approval	
Unanimously approved at the proposed biosafety levels	

AMENDMENT FOR IBC REVIEW

PI	**Saphire**
Protocol #	**BHR16-ES**
Title	Monoclonal antibody neutralization of measles virus
Experimental Procedures	
Agent	Recombinant Measles virus strain B3 encoding eGFP
Amendment summary (from form)	We would like to evaluate the potential for virus evolutionary escape in the presence of a single mAb or cocktails of 2 mAbs in vitro. This would require us to serially passage virus in Vero cells in the presence of the mAbs, then collect and sequence progeny viruses. Viruses would be collected and lysed for RNA extraction in the Saphire lab BSL2 biosafety cabinets using BSL3 safety practices to contain aerosolized virus particles.
Additional details from the protocol	Biosafety Note: The new experiments will not change the biosafety level. R0 is 16-18
Manipulations planned	Same as on the original protocol with the addition of mRNA preparation for sequencing
Recombinant or Synthetic Nucleic Acids	
Source of nucleic sequences (e.g., species)	Measles virus, jelly fish
Nature of nucleic acid (NA) sequences (e.g., enzyme, oncogene)	Genome and marker
NA Host(s) and Vector(s)	Human cell lines
Risk Assessment/Training	
Proposed Risk Assessment	Medium
Training	Verified and on record
IBC Assessment	
Assigned Biosafety Level	BSL-2 with BSL-3 practices aimed at containing the aerosols
CA ATP-L	Yes

NIH Guidelines	III-D-3-a
Category 1 Research	No
Category 2 Research	No
IBC Approval	
Unanimously approved at the proposed biosafety levels with the discussed modifications Note: The IBC discussed the proposed amendment and agree that this was not category 2 research.	

AMENDMENTS APPROVED BY BIOSAFETY

5 protocols amendments were submitted and will be approved by Biosafety (BHR06-BP, BHR03-SC, BHR01-SS, BHR08-SS, BHR01-PV).

These protocol amendments were submitted between August 14th and October 8th. No significant changes were made to the protocols, except for changes related to personnel, funding source, IRB number, IACUC protocol number, or addition of genes, strains or experimental procedures not affecting biosafety level. These minor changes will be approved administratively by the EH&S office.

ANNUAL MONITORING

14 protocols due for annual monitoring (13 received, 1 closed).

These protocols are due for annual monitoring between September 1, 2025, and October 31, 2025. No significant changes were made to the protocols, except for changes related to personnel, funding source, IRB number, IACUC protocol number, or addition of genes, strains or experimental procedures not affecting biosafety level. These minor changes will be approved administratively by the EH&S office.

CLOSED PROTOCOLS

1 protocol(s) due for annual monitoring was closed (BHR06-MC).

STORAGE MEMO

None

GENERAL BUSINESS

TPS UPDATES
The IBC approved the integration of the new RDR tab within the BHR.

BSL-3 INCIDENTS
None

NIH REPORTABLE INCIDENTS
None

DURC
None

Meeting adjourned at 10:46 am