

# Institutional Biosafety Committee Meeting Minutes

**La Jolla Institute**  
FOR IMMUNOLOGY

**Life Without Disease.**

December 10th, 2025, meeting

A regular meeting of the Institutional Biosafety Committee of the La Jolla Institute for Immunology was held in person on Wednesday, December 10th, 2025, at 9:30 AM, with the option to join via Zoom teleconference.

**The meeting started at 9:30 AM.**

**IBC: 12 MEMBERS AND 1 ALTERNATE**

**ATTENDANCE: 8 VOTING MEMBERS, 1 ALTERNATE, 7 MEMBERS REQUIRED FOR QUORUM**

<b>Regular Members</b>	<b>Present</b>
Miguel Reina-Campos, Ph.D. (Chair)	<input checked="" type="checkbox"/>
Mike Barajas (Alternate) CMAR, RLATG	<input checked="" type="checkbox"/>
Sylvie Blondelle, Ph.D.	<input checked="" type="checkbox"/>
Laurence Cagnon, Ph.D.	<input checked="" type="checkbox"/>
Beth Ford, D.V.M.	<input checked="" type="checkbox"/>
David Hall, CSP	<input checked="" type="checkbox"/>
Peter Jones, BS, LATG	<input checked="" type="checkbox"/>
Alessandro Sette, Ph.D.	<input type="checkbox"/>
Stephen Schoenberger, Ph.D.	<input type="checkbox"/>
Kristine Suchey, BS, RVT	<input checked="" type="checkbox"/>
Renna Wolfe, Ph.D.	<input type="checkbox"/>
Jeremy Young, BS, MBA	<input type="checkbox"/>
Marianne Zupanc, Ph.D.	<input checked="" type="checkbox"/>
<b>Others Present:</b> Rose Engel, Hayley Simon	

Note: Stephen Schoenberger attempted to join via zoom.

Introduction of Rose Engel who will replace Jeremy Young as voting member on the IBC. Jeremy will become an alternate member.

## REVIEW AND APPROVAL OF THE MINUTES

The October meeting minutes were unanimously approved

**PROTOCOL REVIEW**

**NEW PROTOCOLS**

<b>PI</b>	<b>Palmer</b>
<b>Protocol #</b>	<b>BHR01-CP</b>
<b>Title</b>	Antisense proof of concept
<b>Experimental Procedures</b>	
<b>Agent</b>	Chemically modified oligonucleotides 18 to 22 bases in length
<b>Project summary (from form)</b>	We want to evaluate efficacy and safety of chemically modified oligonucleotides (18 to 22 nt) targeting specific genes.
<b>Additional details from the protocol</b>	N/A
<b>Manipulations planned</b>	In vivo injections
<b>Recombinant or Synthetic Nucleic Acids</b>	
<b>Source of nucleic sequences (e.g., species)</b>	Synthetic
<b>Nature of nucleic acid (NA) sequences (e.g., enzyme, oncogene)</b>	Antisense (downregulators)
<b>NA Host(s) and Vector(s)</b>	rodents
<b>Risk Assessment/Training</b>	
<b>Risk Assessment</b>	Low
<b>Training</b>	Verified and on record
<b>IBC Assessment</b>	
<b>Proposed Biosafety Level</b>	ABSL-1
<b>CA ATP-L</b>	No
<b>NIH Guidelines</b>	Exempt (III-F)
<b>Category 1 Research</b>	No
<b>Category 2 Research</b>	No
<b>IBC Approval</b>	
Unanimously approved at the proposed biosafety levels with the discussed modifications	

<b>PI</b>	<b>Song</b>
<b>Protocol #</b>	<b>BHR21-KKNA</b>
<b>Title</b>	Adenoviral Vectors
<b>Experimental Procedures</b>	
<b>Agent</b>	Adenoviral vectors
<b>Project summary (from form)</b>	The purpose of this study is to evaluate in vivo hematopoietic stem and progenitor cell (HSPC) mobilization and transduction efficiency following administration of the investigational Adenoviral Vector. Pharmacologic mobilization is required to transiently increase circulating HSPCs, thereby enabling efficient in vivo gene delivery and downstream assessment of engraftment, hematopoietic output, and functional correction.
<b>Additional details from the protocol</b>	N/A
<b>Manipulations planned</b>	In vivo injections
<b>Recombinant or Synthetic Nucleic Acids</b>	
<b>Source of nucleic sequences (e.g., species)</b>	Adenovirus 5, Jelly fish, mouse
<b>Nature of nucleic acid (NA) sequences (e.g., enzyme, oncogene)</b>	Viral vector, marker, enzyme

<b>NA Host(s) and Vector(s)</b>	rodents
<b>Risk Assessment/Training</b>	
<b>Risk Assessment</b>	Low
<b>Training</b>	Verified and on record
<b>IBC Assessment</b>	
<b>Proposed Biosafety Level</b>	BSL-2 and ABSL-2
<b>CA ATP-L</b>	No
<b>NIH Guidelines</b>	III-D-1-a and III-D-4-b
<b>Category 1 Research</b>	No
<b>Category 2 Research</b>	No
<b>IBC Approval</b>	
Unanimously approved at the proposed biosafety levels with the discussed modifications	

**RENEWALS**

<b>PI</b>	<b>Benedict</b>
<b>Protocol #</b>	<b>BHR05-CB</b>
<b>Title</b>	Herpes Virus Simplex Virus - 1 (HSV-1)
<b>Experimental Procedures</b>	
<b>Agent</b>	HSV (wt and recombinant for GFP)
<b>Project summary (from form)</b>	The goal of the project is to perform the screening of different small molecules and assess their capacity to inhibit entry and spread of infection of the Herpes Simplex Virus - 1 (HSV-1) into human cell lines (in vitro). By doing this, it's possible to propose new and more effective treatments and vaccine candidates for HSV-1.
<b>Additional details from the protocol</b>	N/A
<b>Manipulations planned</b>	Tissue culture, viral production, microscopy on fixed or unfixed plates of cells (96 or 384 wells): Centrifuging, pipetting, mixing, plate washing
<b>Recombinant or Synthetic Nucleic Acids</b>	
<b>Source of nucleic acid sequences (e.g., species)</b>	HSV, Jelly fish
<b>Nature of nucleic acid (NA) sequences (e.g., enzyme, oncogene)</b>	Viral genome, marker
<b>NA Host(s) and Vector(s)</b>	Human cell lines
<b>Risk Assessment/Training</b>	
<b>Risk Assessment</b>	Low
<b>Training</b>	Verified and on record
<b>IBC Assessment</b>	
<b>Approved Biosafety Level</b>	BSL-2 with BSL-3 practices aimed at containing the aerosols
<b>CA ATP-L</b>	Yes
<b>NIH Guidelines</b>	III-D-1-a
<b>Category 1 Research</b>	No
<b>Category 2 Research</b>	No
<b>IBC Approval</b>	
Unanimously approved at the proposed biosafety levels with the discussed modifications	

<b>PI</b>	<b>Cheroutre</b>
<b>Protocol #</b>	<b>BHR07-HC</b>
<b>Title</b>	Inducing OVA specific T cell responses in the small intestine. (Salmonella)
<b>Experimental Procedures</b>	
<b>Agent</b>	Salmonella enterica serovar typhimurium (STM) strains include 1) a wild-type, Nalidixic Acid resistant derivative of ATCC 14028 (referred to as Salmonella-IR715) and 2) a strain that expresses chicken egg albumin (OVA) (referred to as Salmonella-OVA)
<b>Project summary (from form)</b>	We want to test if food antigens are important in protecting against intestinal infection.
<b>Additional details from the protocol</b>	N/A
<b>Manipulations planned</b>	Bacteria growth, in vivo infections
<b>Recombinant or Synthetic Nucleic Acids</b>	
<b>Source of nucleic sequences (e.g., species)</b>	HSV, Jelly fish
<b>Nature of nucleic acid (NA) sequences (e.g., enzyme, oncogene)</b>	Viral genome, marker
<b>NA Host(s) and Vector(s)</b>	Rodents
<b>Risk Assessment/Training</b>	
<b>Risk Assessment</b>	Low
<b>Training</b>	Verified and on record
<b>IBC Assessment</b>	
<b>Approved Biosafety Level</b>	BSL-2 and ABSL-2
<b>CA ATP-L</b>	No
<b>NIH Guidelines</b>	III-D-1-a and III-D-4-b
<b>Category 1 Research</b>	No
<b>Category 2 Research</b>	No
<b>IBC Approval</b>	
Unanimously approved at the proposed biosafety levels with the discussed modifications	

<b>PI</b>	<b>Shresta</b>
<b>Protocol #</b>	<b>RDR08-SS</b>
<b>Title</b>	DNA vaccine for flavivirus vaccination
<b>Experimental Procedures</b>	
<b>Agent</b>	DNA plasmids expressing viral proteins
<b>Project summary (from form)</b>	The goal of this project is to develop and investigate the efficacy and mechanisms of DNA plasmid vaccines against flavivirus infection including Zika and Dengue. In vivo immunization with DNA plasmids expressing structural (C, prM, E) or nonstructural proteins (NS1, NS2A, NS2B, NS3, NS4A, NS4B, NS5) from Zika or Dengue genome will be conducted. The DNA plasmid vaccines could express viral proteins alone or in combination. The induction of virus-specific immune responses and viral challenges will be investigated to assess the vaccine-mediated protection.
<b>Additional details from the protocol</b>	N/A
<b>Manipulations planned</b>	In vivo injections
<b>Recombinant or Synthetic Nucleic Acids</b>	
<b>Source of nucleic sequences (e.g., species)</b>	Bacterial, flavivirus proteins

<b>Nature of nucleic acid (NA) sequences (e.g., enzyme, oncogene)</b>	Expression plasmid, structural or non-structural viral proteins
<b>NA Host(s) and Vector(s)</b>	Rodents
<b>Risk Assessment/Training</b>	
<b>Risk Assessment</b>	Low
<b>Training</b>	Verified and on record
<b>IBC Assessment</b>	
<b>Approved Biosafety Level</b>	ABSL-1
<b>CA ATP-L</b>	No
<b>NIH Guidelines</b>	III-D-2-a and III-D-4-a
<b>Category 1 Research</b>	No
<b>Category 2 Research</b>	No
<b>IBC Approval</b>	
Unanimously approved at the proposed biosafety levels	

**AMENDMENT FOR IBC REVIEW**

<b>PI</b>	<b>Saphire</b>
<b>Protocol #</b>	<b>BHR17-ES</b>
<b>Title</b>	Mumps Virus
<b>Experimental Procedures</b>	
<b>Agent</b>	WT or Recombinant Mumps virus encoding GFP
<b>Amendment summary (from form)</b>	Updating agent: Adding a recombinant mumps virus that encodes a GFP or mCherry reporter
<b>Additional details from the protocol</b>	Biosafety Note: The new experiments will not change the biosafety level or the assessed risk, but the project is now subject to NIH guidelines.
<b>Manipulations planned</b>	Same as on the original protocol.
<b>Recombinant or Synthetic Nucleic Acids</b>	
<b>Source of nucleic sequences (e.g., species)</b>	Mumps virus, jelly fish or sea anemone
<b>Nature of nucleic acid (NA) sequences (e.g., enzyme, oncogene)</b>	Genome and marker
<b>NA Host(s) and Vector(s)</b>	Human cell lines
<b>Risk Assessment/Training</b>	
<b>Risk Assessment</b>	Medium
<b>Training</b>	Verified and on record
<b>IBC Assessment</b>	
<b>Approved Biosafety Level</b>	BSL-2 with BSL-3 practices aimed at containing the aerosols
<b>CA ATP-L</b>	Yes
<b>NIH Guidelines</b>	III-D-1-a and III-D-3-a
<b>Category 1 Research</b>	No
<b>Category 2 Research</b>	No
<b>IBC Approval</b>	
Unanimously approved at the proposed biosafety levels with the discussed modifications	

**AMENDMENTS APPROVED BY BIOSAFETY**

11 protocols amendments were submitted and approved by the Biosafety Office (BHR03-SC, BHR01-AG, BHR06-BP, BHR03-BP, BHR01-ES, BHR14-ES, RDR13-ES, BHR01-SS, BHR01-MRC, RDR01-MRC, BHR05-MRC)

*IBC Meeting Agenda Continued*

These protocol amendments were submitted between October 9 and December 9, 2025. No significant changes were made to the protocols, except for changes related to personnel, funding source, IRB number, IACUC protocol number, or addition of genes, strains or experimental procedures not affecting biosafety level. These minor changes will be approved administratively by the EH&S office.

**ANNUAL MONITORING**

**19 protocols due for annual monitoring (15 received, 2 closed, 2 due).**

These protocols are due for annual monitoring between November 1, 2025, and December 31, 2025. No significant changes were made to the protocols, except for changes related to personnel, funding source, IRB number, IACUC protocol number, or addition of genes, strains or experimental procedures not affecting biosafety level. These minor changes will be approved administratively by the EH&S office.

**CLOSED PROTOCOLS**

**2 protocols due for annual monitoring were closed (BHR20-KKNA and RDR05-KKNA).**

**STORAGE MEMO**

None

**GENERAL BUSINESS**

**TPS UPDATES**

Requested comments on the new RDR tab on the BHR form

**BSL-3 INCIDENTS**

None

**NIH REPORTABLE INCIDENTS**

None

**DURC**

None

**Meeting adjourned at 10:12 am**